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Description

This invention relates to processes for the production of immunoglobulin molecules or immunologically functional Ig fragments in cells of host organisms which have been transformed by recombinant DNA techniques.

In recent years advances in molecular biology based on recombinant DNA techniques have provided processes for the production of heterologous (foreign) polypeptides or proteins in host cells which have been transformed with heterologous DNA sequences which code for the production of these products.

Theoretically, the recombinant DNA approach may be applied to the production of any heterologous polypeptide or protein in a suitable host cell, provided that appropriate DNA coding sequences can be identified and used to transform the host cell. In practice, when the recombinant DNA approach was first applied to the production of commercially useful products, its application for the production of any specified polypeptide or protein presented particular problems and difficulties, and the success of applying this approach to the production of any particular polypeptide or product was not readily predictable.

However, a large number of heterologous single chain polypeptides or proteins have now been produced by host cells transformed by recombinant DNA techniques. Examples of such heterologous single chain polypeptides or proteins include human interferons, the A and B chains of human insulin, human and bovine growth hormone, somatostatin, calf prochymosin and urokinase. Such transformed host cells provide a reproducible supply of authentic heterologous polypeptide or protein which may be produced on an industrial scale using industrial fermentation technology.

It should be pointed out that some of these polypeptides, for instance urokinase, after secretion by a host cell appear as two chain molecules. However, in such cases, the molecule is synthesised by the host cell as a single chain polypeptide, coded for by a single DNA sequence, which is cleaved in the host cell subsequent to synthesis to form the two chain structure.

It is known that in both human and animal systems there are a number of polypeptides or proteins which have multichain structure in which the chains are not derived from the cleavage of a single chain polypeptide coded for by a single DNA sequence. In such cases, the gene for each of the chains may be located at different points on the same chromosome or even on different chromosomes. In these cases, the polypeptide chains are synthesised separately and then assembled into the complete molecule subsequent to synthesis. Heretofore, no such multichain polypeptide or protein has been produced by recombinant DNA techniques from a single host cell.

A particular example of a class of such multichain polypeptides or proteins is the immunoglobulins.

Immunoglobulins, commonly referred to as antibodies, are protein molecules produced in animals by B-lymphocyte cells in response to challenge with foreign antigenic agents such as bacteria, viruses and foreign proteins. The immunoglobulins comprise a crucial part of the immune systems of humans and animals. The immunoglobulins recognise specific parts of the foreign agents and bind onto them. The specific parts are usually known as antigenic determinants or antibody binding sites. A given foreign agent is likely to have a number of different antigenic determinants.

A typical immunoglobulin (Ig) molecule is shown in Figure 1 of the accompanying drawings, to which reference is now made. The Ig molecule comprises two identical polypeptide chains of about 600 amino acid residues (usually referred to as the heavy chains, H), disulphide bonded to each other, and two identical shorter polypeptide chains of about 220 amino acid residues (usually referred to as the light chains, L), each light chain being disulphide bonded to one end of each heavy chain as shown.

When the Ig molecule is correctly folded, each chain is formed into a number of distinct globular areas, usually known as domains, joined by a more linear polypeptide chain. The light chains have two such domains, one of which is of variable sequence V_L , and the other of which is of constant sequence C_L . The heavy chains have a single variable domain V_H , adjacent the variable domain V_L of the light chain, and three or four constant domains C_H1-3 or 4.

The variable domains on both the heavy and the light chains each contain three hypervariable regions (HV1-3) which, when the Ig molecule is correctly folded, are located adjacent one another and form the antigen binding site. It is this area which recognises and binds to the antigenic determinant for which the Ig molecule is specific.

The constant domains of the Ig molecule do not take part in the binding to the antigenic determinant, but mediate the actions triggered by the binding of the Ig molecule to the antigenic determinant. It is believed that this triggering is caused by an allosteric effect induced by the binding of the Ig molecule to the antigenic determinant. The constant domain may enable the Ig molecule to fix complement or may cause mast cells to release histamine.

Igs may be categorised by class or subclass, depending on which of a number of possible heavy chain constant domains they contain, there being eight such possible heavy chains in mice. Thus, for instance, an Ig molecule with a μ heavy chain belongs to the class IgM, and one with a γ_1 heavy chain to the class IgG₁.

Igs may also contain one of two light chains, designated as κ and λ light chains, which have different constant domains and different sets of variable domains.

The structure of the Ig molecule and the location of the genes coding for the various domains thereof are discussed more fully by Early and Hood, in *Genetic Engineering, Principles and Methods*, Vol. 3, pages 153-158 (edited by Setlow and Hollaender, Plenum Press).

It is known that Ig molecules on digestion with selected enzymes can produce a number of immunologically functional fragments. Two such fragments are known as the Fab and (Fab')₂ fragments. The Fab fragment comprises one light chain linked to the V_H and C_{H1} domains of a heavy chain as shown in Figure 1. The (Fab')₂ fragment consists essentially of two Fab fragments linked together by a small additional portion of the heavy chains as shown in Figure 1. These fragments and other similar fragments can be of use in various tests and diagnostic and medical methods.

The principle method employed for the production of Igs involves the immunisation of susceptible animals with the antigenic agent to provide an immune reaction. Generally the animal is immunised a second time to improve the yield of Ig. The animal is then bled and the Ig is recovered from the serum.

However, the product of this method is not a homogeneous protein. The animal will produce Igs of different classes and also Igs specific for each of the different antigenic determinants on the antigenic agent, and its blood will therefore contain a heterogeneous mixture of Igs. Obtaining a specific Ig of particular class and desired specificity from such a mixture requires very difficult and tedious purification procedures.

Recently, it has become possible to produce a homogeneous Ig of a single class and a single specificity by a technique first described by Kohler and Milstein (*Nature*, 256, 495-479, 1975). The technique involves the fusion of single Ig-producing parent cells with cancer cells to produce a monoclonal hybridoma cell which produces the Ig. Ig produced by this technique is usually known as monoclonal antibody. The nature of the monoclonal Ig is determined by the class and specificity of the Ig produced by the parent cell.

Recently, attempts have been made to use recombinant DNA techniques to produce fragments of Ig molecules. For instance, Amster *et al.* (*Nucleic Acids Research*, 8, 9, 1980, pp 2055 to 2065) disclose the cloning of double stranded cDNA sequences encoding a mouse Ig light chain into a plasmid. An *E. Coli* strain transformed by the plasmid synthesised a protein thought to comprise the complete constant domain of the light chain and about 40 amino acid residues of its variable region.

Kemp and Cowman (*Proc. Natl. Acad. Sci. USA*, 78, 1981, pp 4520 to 4524) disclose the cloning of cDNA sequences encoding mouse heavy chain fragments and the transforming of an *E. Coli* strain which then synthesised heavy chain polypeptide fragments.

In both these cases, the polypeptides were produced as fusion proteins, in which the fragments of the Ig polypeptides were fused with additional non-Ig polypeptide sequences, and with incomplete variable domains. Thus, the polypeptide chains produced in these studies were not immunologically functional polypeptides as they were incapable of combining with complementary heavy or light chains to provide Ig molecules having intact antigen binding sites and immunological function.

Research studies have also been carried out in mammalian systems. For instance, Falkner and Zachau (*Nature*, 298, 1982, pp 286 to 288) report the cloning of cDNA sequences encoding mouse light chains into a plasmid which was used to transfect genomic eukaryotic cells which could then transiently synthesise light chains.

Rice and Baltimore (*Proc. Natl. Acad. Sci. USA*, 79 1982, pp 7862 to 7865) report on the transfection of a functionally rearranged K light chain Ig gene into a murine leukaemia virus-transformed lymphoid cell line. The cell line is then able to express the gene continuously. In both these cases, the K genes used to transfect the mammalian cells were obtained from myeloma cells and the K polypeptides produced were of indeterminate immunological function.

A further approach is exemplified in a series of papers by Valle *et al.* (*Nature*, 291, 1981, 338-340; *Nature*, 300, 1982, 71-74; and *J. Mol. Biol.*, 160, 1982, 459-474), which describe the microinjection of mRNAs encoding heavy or light chains of Ig isolated from a mouse myeloma line into oocytes of *Xenopus laevis*. Under certain conditions complete Ig molecules were formed. However, the mRNAs were obtained from myeloma cells and the Ig molecules were of indeterminate immunological function.

It can thus be seen that hitherto it has not been possible to produce functional Ig by recombinant DNA technology.

According to a first aspect of the present invention, there is provided a process for producing a heterologous Ig molecule or an immunologically functional Ig fragment in a single host cell, which comprises transforming the host cell with separate DNA sequences respectively encoding polypeptide chains comprising at least the variable domains of the Ig heavy and light chains and expressing each of said polypeptide chains separately in said transformed single host cell.

According to a second aspect of the present invention, there is provided a vector for use in the above process, comprising separate DNA sequences respectively encoding polypeptide chains comprising at least the variable domains of the Ig heavy and light chains.

Preferably, the vector is a plasmid.

The present invention also provides a host cell transformed with a vector as defined above.

The present invention further provides a host cell transformed with two separate vectors, the first vector comprising a DNA sequence encoding a polypeptide chain comprising at least the variable domain of the Ig heavy chain, and the second vector comprising a DNA sequence encoding a polypeptide chain comprising at least the variable domain of the Ig light chain.

In the production of Ig molecules according to the invention it will be appreciated that, in order to produce a functional molecule, the DNA sequences used to transform the host cell will need to encode for at least the V_L and V_H domains of an Ig molecule. Moreover, these domains will need to be complementary so that when the two polypeptide chains fold together they form an antigen binding site of predetermined specificity.

Preferably, the Ig molecule or fragment produced by the process includes a complete light chain and at least the C_{H1} domain in addition to the V_H domain of the heavy chain. Most preferably the process produces a complete Ig molecule.

Advantageously, the Ig molecule or functional fragment thereof produced according to the present invention has a variable region (formed by the V_L and V_H domains) which defines a binding site for an antigenic determinant of clinical or industrial importance. The DNA coding sequences necessary to produce such a molecule may be derived from naturally occurring or hybridoma (monoclonal) Ig-producing cells with the desired specificity.

The constant domains of the Ig molecule or fragment, if present, may be derived from the same cell line as the variable region. However, the constant domains may be specifically altered, partially or completely omitted, or derived from a cell line producing a different class of Ig to provide Ig molecules or fragments having desired properties.

For example, an Ig molecule may be produced having variable domains (V_H and V_L) identical with those from a monoclonal antibody having a desired specificity, and constant domain(s) from a different monoclonal antibody having desired properties, for instance to provide human compatibility or to provide a complement binding site.

Such alterations in the amino acid sequence of the constant domains may be achieved by suitable mutation or partial synthesis and replacement or partial or complete substitution of appropriate regions of the corresponding DNA coding sequences. Substitute constant domain portions may be obtained from compatible recombinant DNA sequences.

The invention may be utilised for the production of Ig molecules or fragments useful for immunopurification, immunoassays, cytochemical labelling and targetting methods, and methods of diagnosis or therapy. For example, the Ig molecule or fragment may bind to a therapeutically active protein such as interferon or a blood clotting factor, for example Factor VIII, and may therefore be used to produce an affinity chromatography medium for use in the immunopurification or assay of the protein.

It is also envisaged that the Ig molecule may be synthesised by a host cell with another peptide moiety attached to one of its constant domains. Such a further peptide moiety may be cytotoxic or enzymatic. Alternatively, the moiety may be useful in attaching the Ig molecule to a biological substrate, such as a cell or tissue, or to a non-biological substrate, such as a chromatography medium. Such a peptide moiety is herein referred to as a structural peptide moiety.

It is further envisaged that cytotoxic, enzymic or structural peptide moieties could be attached to the Ig molecule by normal peptide chemical methods, as are already known in the art, rather than by being synthesised with the Ig molecule.

The Ig molecule or fragment may also comprise a therapeutic agent in its own right. For instance, an Ig molecule or fragment specific for D blood group antigen may be useful for the prevention of haemolytic disease of the new born.

Any suitable recombinant DNA technique may be used in the process of the invention for the production of Ig molecules or fragments. Typical expression vectors such as plasmids are constructed comprising DNA sequences coding for each of the chains of the Ig molecule or fragment.

It will be appreciated that a single vector may be constructed which contains the DNA sequences coding for more than one of the chains. For instance, the DNA sequences coding for Ig heavy and light chains may be inserted at different positions on the same plasmid.

Alternatively, the DNA sequence coding for each chain may be inserted individually into a plasmid, thus producing a number of constructed plasmids, each coding for a particular chain. Preferably the plasmids into which the sequences are inserted are compatible.

The or each constructed plasmid is used to transform a host cell so that each host cell contains DNA sequences coding for each of the chains in the Ig molecule or fragment.

For use in cloning in yeast systems, suitable expression vectors include plasmids based on a 2 micron origin.

Any plasmid containing an appropriate mammalian gene promoter sequence may be used for cloning in mammalian systems. Such vectors include plasmids derived from, for instance, pBR322, bovine papilloma virus, retroviruses, DNA viruses and vaccinia viruses.

Suitable host cells which may be used for expression of the Ig molecule or fragment include yeasts, such as *S. cerevisiae*, and eukaryotic cells, such as insect or mammalian cell lines.

The present invention also includes constructed expression vectors and transformed host cells for use in producing the Ig molecules or fragments.

After expression of the individual chains in the same host cell, they may be recovered to provide the Ig molecule or fragment in active form, for instance to provide an Ig molecule of predetermined immunological function.

It is envisaged that in preferred forms of the invention, the individual chains will be processed by the host cell to form the complete polypeptide or protein which advantageously is secreted therefrom.

It will be appreciated that the present application shows for the first time that it is possible to transform a host cell so that it can express two or more separate polypeptides which may be assembled to form a complete Ig molecule or fragment. There is no disclosure or suggestion of the present invention in the prior art, which relates solely to the production of a single chain heterologous polypeptide or protein from each host cell.

The present invention will now be described, by way of example only, with reference to the accompanying drawings, in which:-

Figure 1 shows a diagrammatic representation of a typical intact Ig molecule;

Figure 2 shows the construction of plasmids for the direct synthesis of a λ light chain in *E. Coli*;

Figure 3 shows the construction of plasmids for the direct synthesis of a μ heavy chain in *E. Coli*;

Figure 4 is a diagrammatic representation of μ mRNA sequences around the initiation codon;

Figure 5 shows the construction of plasmids having altered secondary structure around the initiation codon;

Figure 6 is a polyacrylamide gel showing expression and distribution of μ protein from *E. Coli* B;

Figure 7 is a polyacrylamide gel showing pulse chase autoradiograms of μ protein in *E. Coli* B and in *E. Coli* HB101;

Figure 8 is a polyacrylamide gel showing the results of λ gene expression in *E. Coli*;

Figure 9 is a polyacrylamide gel showing the distribution of recombinant λ light chain polypeptide between the soluble and insoluble cell fractions;

Figure 10 is a polyacrylamide gel showing expression and distribution of λ protein from *E. Coli* E103S;

Figure 11 shows the results of the fractionation of μ and λ protein expressed by *E. Coli* B on DEAE Sephacel;

Figure 12 shows the specific hapten binding of reconstituted Ig molecules; and

Figure 13 shows the heteroclitic nature of the hapten binding of the reconstituted Ig molecules.

In the following examples, there is described the production of Ig light and heavy chain polypeptides derived from monoclonal antibodies which recognise and bind to the antigenic determinant 4-hydroxy-3-nitrophenyl acetyl (NP), using *E. coli* and *S. cerevisiae* as the host cells. Recombinant DNA techniques were used to enable the host cells to express both the polypeptide chains.

It will be appreciated that the invention is not limited to the specific methods and construction described hereafter.

Construction of Lambda Light Chain Expression Plasmid

Figure 2, to which reference is now made, shows schematically the method used to construct a λ light chain expression plasmid.

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R45 5' -pGATCAATGCAGGCTGTTGTG 3'

R44 3' CCGACAACACTGAGTCCTTA_p- 5'

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The sequence in this area was:

...GATTGATCA.ATG.CAG.GCT.GTT.ATG.ACT.CAG.GAA.TCT.GCA.CTC.ACC.ACA.TCA.

met gln ala val met thr gln glu ser ala leu thr thr ser

50

The plasmids which had lost their *Cla*I site were sequenced in the region surrounding the initiator ATG.

5 AAGGGTATTGATCAATG CAG..... plasmid pNP3
 SD met glu

10 AAGGGTTTGATCAATG CAG plasmid pNP4
 SD met glu

15 In order to achieve high level expression a number of other approaches were followed. Firstly, a series of constructs were obtained which had increasing amounts of the 3' untranslated region of the cDNA removed by *Bal* 31 exonuclease. Secondly, a high copy number plasmid containing λ cDNA was constructed. This plasmid contained a par function (Meacock, P. A. and Cohen, S.N. Cell, 20, 529-542, 1980) as well as being present in high copy number. Thirdly, the pNP3 plasmid was transformed into a number of protease-deficient strains or into HB101 in conjunction with a protease deficient dominant acting plasmid (Grossman, A.D. et al Cell, 32, 151-159, 1983).

20 Construction of μ Heavy Chain Expression Plasmid

The full-length μ heavy chain cDNA derived from the NP binding monoclonal antibody B1-8 had been cloned into the *Pst*I site of pBR322 yielding a plasmid designated pAB μ -11 (Bothwell et al, Cell, 24, 625-637, 1981). In order to achieve high level expression, the μ cDNA minus the eukaryotic leader was reconstructed into pCT54. The construction of the μ heavy chain expression plasmid is shown diagrammatically in Figure 3. Two chemically synthesised oligonucleotides were made to facilitate this. These have the following sequences:

30 R43 5' GATCAATGCAGGTTTCAGCTGCA 3'
 R46 3' TTACGTCCAAGTCG 5'

35 These were ligated into *Bcl*II cut pCT54 using T4 DNA ligase and the resulting plasmid designated pCT54 Pst. The linkers were designed to replace the sequence in pCT54 between the *Bcl*II site and the ATG, to provide an internal *Pst*I site and to recreate the sequence from pAB μ -11 5' to the *Pst*I site up to codon +1. pCT54 Pst was cut briefly with *Pst*I, treated with alkaline phosphatase and full length DNA glass isolated from a 1% agarose gel. Similarly pAB μ -11 was briefly cut with *Pst*I and the full length μ insert isolated following agarose gel electrophoresis. The μ cDNA was ligated with the full length pCT54Pst fragment using T4 DNA ligase under standard conditions and a plasmid designated pCT54 μ was identified which by restriction enzyme analysis was shown to contain a full-length μ insert. The plasmid was sequenced around the 5' linker region and was found to have the anticipated sequence:

45 5'TGATCAATGCAGGTTTCAGCTGCAGGGGGGGGATGGGATGGAG.... 3', demonstrating that it was, indeed, a full length clone. A complete *Pst*I digest of pCT54 μ liberated a 1.4 Kb fragment which was purified by 0.8% agarose gel electrophoresis and glass powder isolation. This was ligated with *Pst*I cut pCT54 Pst (see above) using T4 DNA ligase under standard conditions, followed by transformation into HB101. A plasmid designated pNP1 was isolated which was shown by restriction endonuclease pattern analysis to contain the 1.4 Kb μ cDNA fragment in an appropriate orientation (Figure 3). pNP1 was a plasmid which consisted of an appropriate 5' end for expression, whilst pCT54 contained an appropriate 3' end. The full length gene was reconstructed into pCT54 by cutting both pNP1 and pCT54 μ with *Ava*I which cuts once in the plasmid and once in the μ gene. Both digests were run on a 1% agarose gel and the 1.9 Kb fragment from pNP1 and the 3.65 Kb fragment from pCT54 μ isolated. Following alkaline phosphatase treatment of the 3.65 Kb pCT54 μ fragment, the two pieces of DNA were ligated to each other and transformed into HB101. A plasmid designated pNP2 was identified which demonstrated the correct restriction endonuclease pattern. It was sequenced in the area surrounding the initiator ATG and found to have the anticipated sequence:

5'TTGATCAATGCAGGTTTCAGCTGCAGCAGCCTGGGGCTGAGCTTGTGAAG.... 3'

The vector pCT54 had been constructed to include two restriction sites (Bcl1 and Cla1) between the S-D sequence (AAGG) and the initiation codon so that the distance between these sequence elements could be varied. As most *E.coli* mRNAs have 6-11 nucleotides between the S-D sequence and the AUG the distance in pNP2 was reduced by modification at the Cla1 site. pNP2 was cut with Cla1 and incubated with S1. The amount of S1 nuclease was adjusted so that some DNA molecules would lose 1-2 extra base pairs as a result of 'nibbling' by the enzyme. This DNA on religation with T4 DNA ligase and transformation into *E.coli* strain HB101 gave rise to a number of colonies harbouring plasmids which had lost the Cla1 site. The sequences around the initiation codon of four plasmids pNP223, pNP261, pNP9 and pNP282 were determined and are given in Table 1, part A below.

TABLE 1

		S-D to ATG distance (no. of residues)	Sequence (5' - 3') Cla1	Expression: (mu) (ru)		
				A	B	C
(A)	pNP2	14	-AAAAGGGTATCGATTGATCAATGAGGTTTCAG-	0.7	0.6	0.8
S1 derivatives of pNP2	pNP223	12	-AAAAGGGTATATTGATCAATGAGGTTTCAG-	-	0.7	0.6
	pNP261	10	-AAAAGGGTATTGATCAATGAGGTTTCAG-	-	1.1	1.1
	pNP282	7	-AAAAGGGTGATCAATGAGGTTTCAG-	-	0.7	0.6
	pNP9	9	-AAAAGGGTATGATCAATGAGGTTTCAG-	1.0	1.0	1.0
(B)	pNP11	11	-AAAAGGGTATTGCACATATGCAAGTGCAA-	5.3	6.3	9.1
oligo constructs	pNP12	11	-AAAAGGGTATTGCACATATGAGGTTTCAG-	1.8	2.3	1.3
	pNP14	9	-AAAAGGGTATGATCAATGCAAGTGCAA-	72.2	106.7	124.6
	pNP8	-	-	77.0	53.4	145.5
	pCT70	-	-	0	0	0

The expression of these constructs in *E. coli* B was examined in three experiments, and concentrations of μ protein were determined by ELISA as relative units (ru), a measure of μ concentration normalised to pNP9. The results are shown in Table 1.

Inductions were carried out by resuspending cells at 1:50 dilution from overnight cultures into induction medium, consisting of: KH_2PO_4 (3g/L), Na_2HPO_4 (6g/L), NaCl (0.5g/L), Difco vitamin assay casamino acids (30g/L), NH_4Cl (4g/L) glycerol (16g/L), proline (0.5g/L), Difco yeast extract (1g/L), $\text{CaCl}_2 \cdot 6\text{H}_2\text{O}$ (0.022g/L), $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ (0.025g/L), thiamine (0.01g/L) and carbenicillin (0.1g/L). Cultures were shaken at 37°C and cells harvested by centrifugation.

ELISA assays were performed essentially as described by Patel et al. (Nucleic Acid Research, 10, 5605 to 5620, 1982), using affinity purified goat anti-mouse IgM (Tago) and affinity purified goat anti-mouse IgM (Tago) conjugated to peroxidase, with 3, 3', 5, 5' tetramethylbenzidine (Miles) as a substrate. Bacterial cell pellets were boiled for 2 minutes in 9M urea (BRL) and loaded directly into the ELISA, such that all wells contained 2.25M urea, 10mM Tris-HCl pH7.8, 150mM NaCl, 0.05% NP40 and 0.5% casein (Hammarsten, B.D.H.).

Altering the S-D to ATG distance with nuclease S1 was found to increase the level of μ expression, relative to the parental plasmid pNP2, but only to a small extent (compare pNP9). The optimal S-D to AUG distance was found to be 9-10 nucleotides, as found in pNP9 and pNP261 respectively.

Secondary Structure Analysis

Hairpin loops were identified by use of the computer programme HAIRGU developed by Dr. Roger Staden. The ΔG values were calculated as described by Trioco et al. (Nature NB, 246, 40-41, 1973).

5 Analysis of the potential secondary structure of μ mRNA encoded by pNP2 and pNP9 (one of the S1 derivatives or pNP2) revealed an extensive array of possible hairpin loops. Attention was focused on the region of this mRNA containing the ribosome binding site since the sequestering of S-D sequences and initiation codons into secondary structure may inhibit the initiation of translation. It was found that a hairpin loop including the U and G of the initiation codon could be formed (see Figure 4). This hairpin loop is
10 formed entirely within coding sequences and has a $\Delta G = -7.6$ kcal. No secondary structures were found which buried the S-D sequence, and none which mutually excluded the hairpin loop described above, and which had a lower ΔG .

To test whether expression of μ protein could be influenced by changes designed to alter potential secondary structure in the ribosome binding region we used synthetic oligonucleotides to mutagenise this
15 region of μ mRNA. The plasmids were constructed by insertion of a pair of oligonucleotides between the blunted Cla I site and the 5' Pst site of μ in pNP2 (see Figure 5). The cloning strategy used to accomplish this, however, was multi-staged due to the presence of four Pst sites in pNP2. The vector pACYC184CM, which lacks Cla I and Pst I sites, was first constructed by cutting pACYC184 at its unique Cla I site, 'filling-in' the cohesive termini using T4 DNA polymerase and religating. Next, pACYC184CM was cut with BamHI,
20 and the BamHI fragment from pNP2, bearing the 5' μ sequence, was inserted into it to form pCM μ (see Figure 4). The vector pCM μ contained unique Cla I and Pst sites which were required for the oligonucleotide cloning (see Figure 5).

pCM μ was cut with Cla I to completion, blunted via nuclease S1, and then cut with Pst I. The cut plasmid was purified by agarose gel electrophoresis and ligated separately with pairs of oligonucleotides,
25 with the latter in a 100-fold molar excess. The pairs of oligonucleotides used were: R131 and R132 (for pNP11); R196 and R197 (for pNP12); and R202 and R203 (for pNP14). These have the sequences given below.

30 TGCACATATGCAAGTGCAACTGCA (R131), GTTGCACCTGCATATGTGCA (R132),
GCTGAACCTGCATATGTGCA (R196), TGCACATATGCAGGTTTCAGCTGCA (R197),
GTTGCACCTGCATTGATC (R202), GATCAATGCAAGTGCAACTGCA (R203).

35 The ligation mixture was then used to transform HB101. Recombinant clones were detected by colony hybridisation, on nitrocellulose filters, using one of each pair of oligonucleotides as a probe. Positive clones were sequenced and shown to have the correct nucleotide sequence. The μ sequences containing the cloned oligonucleotides were excised on Eco RV-3gl II fragments. These were ligated to Eco RV-Bgl II cut
40 pNP2 to reconstruct the full length μ genes (see Figure 5). Three different plasmids were created by these procedures; pNP11, pNP12 and pNP14, having the nucleotide sequences shown in Table 1 above.

In the construct pNP11 the potentially deleterious hairpin loop of pNP2 was abolished by changing the residues in the degenerate position of three codons and by introducing a different A-D to AUG sequence. These changes allow the region between the S-D and AUG to base pair with the 5' end of μ coding
45 sequence to form a hairpin loop of $\Delta G = -7.8$ kcal, approximately equal to that of pNP2, but leaving the AUG and S-D sequences exposed (see Figure 4). The only other mutually exclusive hairpin loops are of $\Delta G = -2.3, -3.6$ and $+0.4$ kcal.

The construct pNP12 retains the S-D to AUG sequence of pNP11, but has the coding sequence of pNP2, such that the hairpin loop of pNP2 can form (see Figure 4).

50 The final μ construct was pNP11, which had the S-D to AUG sequence of pNP9, but the 5' coding sequence of pNP11, so that the hairpin loop of pNP12 could not form. The construct pNP14 had no secondary structure burying the S-D or μ AUG.

μ Protein Expression from Constructs with Differing mRNA Secondary Structures

55 E.coli B strains containing plasmids pNP9, pNP11, pNP12 and pNP14 were grown in induction medium and samples removed for ELISA assay. The concentration of μ was increased 6-7 fold in E.coli B cells containing pNP11 relative to pNP9 (see Table 1, Part b). Plasmid pNP12, which has the S-D to AUG

sequence of pNP11, but the coding sequence of pNP9, showed only a two-fold increase in level of μ expression compared to pNP9.

Plasmid pNP14, which has the S-D to AUG sequence of pNP9 but the 5' μ coding sequence of pNP11, and thus differed from pNP9 only in three residues, was found to express μ protein over 90 times that found in pNP9.

A plasmid, pNP8, has also been constructed, from which μ is expressed, also from the *trp* promoter, as part of a *trpE*- μ fusion protein containing the amino terminal 53 amino acids of *trpE* and the carboxyl terminal 503 amino acids of μ . The rationale for making this fusion gene was that μ would be translated from an efficient bacterial ribosome binding site rather than from one of unknown efficiency. Then, by comparing the amounts of *trpE*- μ produced from pNP8 with those of μ produced from the different constructs, an estimate of relative ribosome binding efficiency may be made. When the *trpE*- μ gene in pNP8 was induced and the products quantitated by ELISA, it was found that the μ expression in strains containing pNP8 and pNP14 were very similar (see Table 1, part B).

The ribosome binding site (RBS) sequences of pNP11, pNP12 and pNP14 were designed in order to test the hypothesis that sequestering of the initiation codon into secondary structure inhibits translation initiation, and to increase the rate of initiation by releasing the initiation codon. In pNP11, the RBS hairpin loop of pNP2 and pNP9 was abolished by changing the 5' μ coding sequence in the degenerate position of three codons. In addition, the S-D to AUG sequence was changed to allow the base-pairing of this region with the 5' coding sequence. The result of these changes is that the initiation codon is exposed in the loop of a hairpin with the S-D sequence at the base of the stem (see Figure 4). This hairpin loop had a ΔG approximately equivalent to the ΔG of the pNP9 hairpin loop. Strains containing pNP11 produced 6-7 times as much μ as did those with pNP9. To test whether this was due to the change in S-D to ATG sequence characteristics, rather than to the liberation of the initiation codon, pNP12 was constructed as a control. The latter has the S-D to AUG sequence of pNP11, but retains the 5' μ coding sequence of pNP9, so that the RBS hairpin loop of pNP11 cannot form, but the potentially deleterious one of pNP9 can form. The level of μ expression from pNP12 was found to be increased by only two-fold relative to pNP9. This increase is probably due to an altered S-D to AUG distance, a U immediately 5' to the initiation codon, and the S-D to AUG sequence being G-poor each of which has been shown to be advantageous. It is therefore likely that the increased expression from pNP11 is due to a preferable mRNA secondary structure. The RBS hairpin loop of pNP9 probably acts to inhibit translation while creation of another hairpin loop which exposes the initiation codon is responsible for the increased expression.

The plasmid pNP14 was constructed, such that it had the S-D to AUG sequence of pNP9, but the coding sequence of pNP11. The expression plasmid pNP14, therefore, differed from pNP9 in only three residues, those in the degenerate position of three codons, yet strains containing pNP14 expressed over ninety times more μ than those containing pNP9. This increase in expression cannot be explained on the basis of the three residue changes optimizing codon usage, for they introduce less favoured codons, as judged by the frequency of their occurrence in strongly expressed *E. coli* genes.

The increased expression is best explained by the abolition of the RBS hairpin loop in pNP9. The magnitude of the increases in expression observed, and the few residue changes introduced, makes it difficult for these results to be interpreted in terms of changes in mRNA stability.

The expression of pNP8, which encodes the *trpE*- μ fusion protein from a native *trpE* RBS, was also examined, and found to express at approximately the same level as pNP14. This may indicate that the RBS of pNP14 is equivalent to that of *trpE* in terms of its efficiency in directing translation initiation. The native *trpE* RBS of pNP8 and that of pNP14 were found to have no secondary structures hindering the use of their RBS sequences.

Expression of μ Protein in *E. coli*

E. coli B cells containing the μ expression plasmid pNP11 were grown under inducing conditions and soluble and insoluble extracts prepared, and analysed by SDS-PAGE. A novel band was seen after staining the gel with Coomassie blue in the lane containing proteins from the insoluble fraction (see Figure 6, lane 2). This band was not seen in the negative control lane which contained proteins from the same fraction from cells harbouring pCT70 (see Figure 6, lane 3). The novel band was found to migrate to a position corresponding to a protein of a molecular weight within less than 5% of the actual molecular weight of non-glycosylated μ of 62.5Kd. A duplicate set of lanes were transferred to nitrocellulose, and Western blotted. Alignment of the stained gel and the blot autoradiogram confirms that this novel band is antigenically related to IgM (see Figure 6, lanes 4 and 8). No band was found in extracts from cells containing pCT70 (see Figure 6, lanes 5 and 7). Only a low amount of μ was found in the soluble fraction (see Figure 6, lane

6).

A greatly increased level of expression of μ was found in *E. Coli* B compared to HB101. Pulse chase analysis demonstrated that in *E. Coli* B, a similar level of μ protein was detected after a 60 minute chase (figure 7, lane 3) as was seen after the initial labelling period (Figure 7, lane 1). In HB101, however, very little μ protein could be seen after a 10 minute chase (Figure 7, lane 7), and none after 30 minutes (Figure 7, lane 8), compared to the amount detected after the initial labelling period (Figure 7, lane 5).

For pulse chase analysis, inductions were set up as described above, except that the medium used consisted of: proline (0.3g/L), leucine (0.1g/L), Difco methionine assay medium (5g/L), glucose (60mg/L), thiamine (10mg/L), CaCl_2 (22mg/L), MgSO_4 (0.25g/L) and carbenicillin (0.1g/L). During exponential growth cells were pulse labelled with 30 $\mu\text{Ci/ml}$ $\text{L-}^{35}\text{S}$ methionine for 2 minutes, after which unlabelled methionine (100 $\mu\text{g/ml}$) was added and the incubation continued for the times indicated.

Induced *E. Coli* B cells harbouring pNP14 when examined by phase contrast microscopy were found to contain inclusion bodies.

15 Expression of λ Light Chain in *E. Coli*

A fresh 1 ml overnight culture of *E. Coli* HB101 or RV308 containing the plasmid under study was grown in L Broth (Maniatis et al., Molecular Cloning, Cold Spring Harbor Laboratory, 1982), supplemented with carbenicillin to 100 $\mu\text{g/ml}$.

20 A 1:100 dilution of this culture was made into M9 medium (Maniatis, 1982, op.cit.) supplemented with glucose, vitamin B₁, carbenicillin, leucine and proline and the cultures shaken at 37° for 5-6 hours. At this time ^{35}S -methionine was added for 5-20 minutes. The cells were then harvested by centrifugation, lysed by boiling in 1% SDS for 2 minutes and diluted by addition of a buffer consisting of 2% Triton X-100, 50 mM Tris pH 8, 0.15 M NaCl and 0.1 mM EDTA. Immunoprecipitations were carried out by addition of antisera to aliquots of labelled *E. Coli* extracts and, after incubation at 4° overnight, immune complexes were isolated by binding to *Staphylococcus aureus* fixed cells. The complexes were dissociated by boiling in 60 mM Tris pH 6.8 buffer containing 10% glycerol, 3% SDS and 5% mercaptoethanol and the liberated proteins analysed on 10% or 12.5% acrylamide/SDS gels (Laemmli, UK. Nature, 227, 680-685, 1970). Gels were stained with Coomassie Brilliant blue to visualise the protein bands while labelled proteins were detected on 30 Fuji-RX film by fluorography using 1 M sodium salicylate.

HB101 cells containing pNP3 and other plasmids were grown under inducing conditions to an $\text{OD}_{600} = 0.6$ and the proteins present examined by specific immunoprecipitation substantially as described above. The results obtained are given in Figure 8, in which lanes 1 and 2 respectively were extracts from pCT54-19 and pNP3, immunoprecipitated with normal rabbit serum; lanes 3, 4 and 5 were extracts immunoprecipitated with rabbit anti-lambda serum and represent pNP4 (lane 3), pNP3 (lane 4) and pCT54-19 (lane 5). The position of unlabelled lambda protein from MOPC104E is indicated on the left hand side of Figure 8. All of the plasmids produced a protein which reacted with rabbit anti-mouse λ light chain serum and comigrated with authentic λ light chain from the mouse myeloma MOPC104E. pNP3 however produced the most of this protein (compare lane 4 with lanes 3 and 5 in Figure 8). No such band was detected with control immunoprecipitations using normal rabbit serum (Figure 8, lanes 1 and 2). Studies using brief pulses with ^{35}S -methionine followed by chasing with excess cold methionine indicated that the recombinant λ light chain had a 1/2 life of about 20 minutes.

In vitro transcription/translation (Pratt et al, Nucleic Acids Research, 9, 4459-447, 1981) confirmed that pNP3 coded for a protein which comigrated with authentic λ light chain. This 25 Kd product was synthesised in vitro at a rate comparable to that of β -lactamase indicating that it is synthesised in vivo at a level of 0.5% of total *E. Coli* proteins. This figure is in good agreement with the percentage of recombinant λ product synthesised in vivo (0.4-2%) as determined from the following equation:

$$50 \quad \% \text{ specific CPM} = \frac{100 \times (\text{CPM with anti-}\lambda) - (\text{CPM with normal serum})}{\text{total CPM}}$$

The procedure used to disrupt cells was as follows (Emtage J S et al, Proc. Natl. Acad. Sci., 80, 3671-3715, 1983). *E. Coli* HB101/pNP3 grown under inducing conditions were harvested and resuspended in 0.05 M Tris pH 8, 0.233 M NaCl, 5% glycerol containing 130 g/ml of lysozyme and incubated at 4°C or room temperature for 20 minutes. Sodium deoxycholate was then added to a final concentration of 0.05% and 10 μg of DNAase 1 (from bovine pancreas) was added per g wet wt of *E. Coli*. The solution was incubated at

15°C for 30 minutes by which time the viscosity of the solution had decreased markedly. The extract was centrifuged (at 10,000 x g for 15 minutes for small volumes (1 ml) or 1 hour for larger volumes) to produce a soluble and an insoluble fraction.

For immunoprecipitations, the soluble fraction was diluted in the Triton-containing buffer described above and the insoluble fraction solubilised by boiling in 1% SDS followed by dilution in Triton-containing buffer. HB101 cells containing pNP3 or pNP4 were grown under inducing conditions, pulse labelled with [³⁵S] methionine, separated into soluble and insoluble fractions and analysed by sodium dodecyl sulfate polyacrylamide gel electrophoresis. The results obtained are shown in Figure 9 in which lanes 1 and 3 correspond to the soluble fraction from HB101-pNP3 immunoprecipitated with normal rabbit serum and rabbit anti-lambda serum respectively; lanes 2 and 4 correspond to the insoluble fraction from HB101-pNP3 immunoprecipitated with normal rabbit serum and rabbit anti-lambda serum respectively, lanes 5 and 7 correspond to the soluble fraction from HB101-pNP4 immunoprecipitated with normal rabbit serum and rabbit anti-lambda serum respectively; lane 9 corresponds to the soluble fraction from HB101-pNP3, and lane 10 corresponds to the insoluble fraction from HB101-pNP3. The position of unlabelled lambda protein from MOPC104E is indicated on the left hand side of Figure 9. When such a procedure was carried out using pNP3, the recombinant lambda light chain protein was present in the insoluble fraction (Figure 9, lane 4) rather than the soluble fraction (Figure 9, lane 3). No 25 Kd band comigrating with authentic lambda light chain was present when either the soluble or insoluble fraction were immunoprecipitated using normal rabbit serum (Figure 9, lanes 1 and 2). By inference it is likely that all Ig light chains will fractionate as in this specific example.

In the absence of specific immunoprecipitation a novel protein band was not visible from extracts of HB101 containing pNP3 nor was the lambda protein found to accumulate. However, there was a dramatic difference when pNP3 was induced in the K12 strain E103s. In this strain lambda protein was found to accumulate during induction until the cells reaches stationary phase (figure 10, lanes 3-5) to a level of about 150 times that found in HB101 as determined by an ELISA (enzyme linked immunosorbent assay). These cells were found to contain inclusion bodies which appeared refractile under light microscopy, a phenomenon characteristic of high level expression of foreign proteins. An estimate of the percentage of total E. Coli protein represented by recombinant lambda protein was obtained by separating the proteins by gel electrophoresis, staining them with Coomassie blue and scanning the stained gel with a Joyce-Loebl chromoscan 3. This method showed that lambda was the major protein present (Figure 10, lane 5) and represented 13% of total E. Coli protein. The lambda protein had a half-life of 20 minute in HB101 but accumulated to very high levels in E103s, suggesting that Lambda protein was much more stable in the latter strain. After cell lysis and centrifugation of HB101 or E103s containing pNP3, lambda light chain was detected in the insoluble (Figure 10, lanes 7 and 10) but not in the soluble fractions (Figure 10, lanes 6), as determined by Coomassie blue staining. The identity of the major Coomassie blue stained band as lambda protein was confirmed by Western blot analysis (Figure 10, lanes 8-10). The presence of such immunoreactive bands was specific to pNP3 containing cells. When extracts from cells containing pCT70, a pro-chymosin expressing plasmid, were subjected to the same analysis no bands were detected. This more sensitive technique showed that a small amount of the lambda protein was in the soluble fraction (Figure 10, lane 9). The presence of a number of distinct immunoreactive proteins all smaller than full-length protein were also detected. These may result from proteolytic degradation of lambda protein, from premature termination of transcription or from internal initiation of translation.

Expression of mu and Lambda Polypeptides in the same Bacterial Cell

Each of the Ig mu and lambda genes in expression plasmids were transformed into the same E.Coli cell to direct the synthesis of both Ig mu and lambda polypeptides. In order to overcome plasmid incompatibility and provide a second antibiotic resistance marker, the trp promoter and lambda sequences were excised from pNP3 on a Hind III - Bam HI fragment and inserted into the Hind III - Bam HI fragment of pACYC 184. The resultant plasmid pACYC lambda caused the E.Coli to grow very poorly. This weak growth was thought to result from read-through of RNA polymerase into the origin of replication. However, the inhibition of growth was virtually eliminated by the cloning in of the bacteriophage T7 transcriptional terminator at the Hind III site of plasmid pACYC lambda.

This terminator functions in both orientations. The resultant plasmid pAC lambda T7-1 has a chloramphenicol resistance gene and an origin compatible with the pBR 322-derived origin on pNP14, the Ig mu expressing plasmid. Transformation of both plasmids in the same E.Coli B was achieved in two steps. Firstly pNP14 was introduced followed by pAC lambda T7 - 1 in two sequential transformations to give ampicillin and chloramphenicol resistant clones.

E.Coli B cells derived from this double-transformant clone showed the presence of inclusion bodies and two novel polypeptide bands on stained gels of the insoluble fraction after lysis. These two bands correspond both with immunological activity by Western blotting for Ig μ and Ig λ and in their expected molecular weights. It has thus been shown that the double-transformant clone expresses both the heterologous genes. This had not hitherto been shown.

The presence of λ light chain in the insoluble fraction was a useful purification step since it both concentrated the protein and separated it from the bulk of E. Coli soluble proteins.

For further purification of the λ light chain, the cell debris were dissolved in 10mM Tris-HCl pH8.0, 25% formamide, 7M urea, 1mM EDTA and 2mM dithiothreitol. This material was loaded onto a DEAE Sephacel column (Pharmacia) (1x 25cm at a flow rate of 5ml/hr) which had been equilibrated in 9M urea, 10mM Tris-HCl pH8.0, 1mM EDTA and 2mM DTT. The DEAE Sephacel column was developed using a 0-150mM NaCl gradient in loading buffer. The eluted peak λ light chain immunoreactivity, corresponding to the major peak of protein, was diluted to a final concentration of 2.25M urea, 10mM Tris-HCl pH8.0, 1mM EDTA, 2mM DTT and loaded onto an octyl-Sepharose column (Pharmacia) (2.5 x 10cm). Material was eluted by use of a urea gradient of 2.25-9M urea. The peak material was pooled, dialysed into ammonium bicarbonate and lyophilised. Following this step, only a single band of Coomassie blue stainable material corresponding to recombinant λ protein was visualised by SDS-PAGE.

The μ heavy chain was purified from 9M urea solubilised pellets by anion exchange chromatography and chromatofocussing (Pharmacia).

It was of great interest to determine whether the concomitant expression of μ and λ would lead to the formation of functional IgM. In order to determine this, extracts were made from E. Coli containing both Ig μ and λ polypeptides and these tested for antigen binding. We used a two-site sandwich ELISA which detects μ chain binding to haptenalated bovine serum albumin (NIP-caproate-BSA).

In the NIP binding assay, bovine serum albumin (BSA) is reacted with an equimolar amount of NIP-Cap-N-hydroxy-succinamide at pH 7.5 in 10mM phosphate buffer. The resulting NIP-Cap-BSA is separated from free NIP-Cap on a G-50 Sephadex column.

Microtitre plates (96 well Nunc Immuno Plate 1) are coated with 100 μ l of a solution of 10 μ g/ml NIP-BSA in-sodium carbonate/sodium bicarbonate 0.1M, pH 9.6 buffer (coating buffer) overnight at 4°C.

The coated plates are then blocked for non-specific binding by addition of 100 μ l of 0.5% casein in coating buffer (blocking buffer) and incubating at 37°C for 1 hour.

The coated and blocked plates are then washed three times in 150 mM NaCl, 0.05% NP40, 20mM Tris-HCl, pH 7.8 buffer (washing buffer).

The washed plates are shaken free of excess washing buffer and samples are added in their buffer or typically in 0.5% casein in washing buffer (sample buffer) to a 100 μ l final volume.

In order to demonstrate the nature of the binding to NIP-BSA, samples for testing were supplemented with either NIP or NP in free solution at various concentrations (from 60 to 0.3 μ M for NIP or from 600 to 3 μ M for NP).

The plates with samples are then incubated at 37°C for one hour and thereafter washed three times with washing buffer.

The washed plates are then inoculated with 100 μ l of anti μ -peroxidase conjugate (1:1000 dilution; TAGO Inc) in sample buffer, and incubated for one hour at 37°C. The plates are then washed three times in washing buffer.

The washed plates are inoculated with 100 μ l of 0.1M Na acetate-citrate, pH 6.0, 0.1 mg/ml tetramethylbenzidine, 13mM H₂O₂ (peroxidase substrate) and incubated at room temperature for one hour.

The reaction is terminated by the addition of 25 μ l of 2.5M H₂SO₄.

The microtitre plates are then read in a plate reader (Dynatech) at 450 nm with a reference of 630 nm. The A_{450} was related to the level of a standard protein, B1-8 anti-NIP IgM.

This assay demonstrates sensitivity to 60pg of B1-8 IgM. The extracts were prepared as soluble and insoluble material. The insoluble material was solubilized in the same buffer used in lysis but containing 8M urea followed by its dilution for assay.

In order to obtain activity for the Ig μ and λ , extracts were made of the insoluble fraction and these dialysed into buffer conditions in which disulphide interchange will occur at a higher frequency.

Production of functional antibodies from E. Coli expressing both heavy and light chains was achieved by lysing the cells and clarifying the supernatant by centrifugation. The insoluble material was washed, followed by sonication (3 times for 3 minutes), and finally dissolved in 9M urea, 50mM glycine-NaOH pH10.8, 1mM EDTA, and 20mM 2-mercaptoethanol. This extract was dialysed for 40 hours against 3 changes of 20 vols. of 100mM KCl, 50mM glycine-NaOH pH10.8, 5% glycerol, 0.05mM EDTA, 0.5mM reduced glutathione and 0.1mM oxidised glutathione. The dialysate was cleared by centrifugation at 30,000g

for 15 minutes and loaded directly onto DEAE Sephacel, followed by development with a 0-0.5M KCl linear gradient in 10mM Tris-HCl, 0.5mM EDTA, pH8.0.

The purified Ig μ and λ were treated as above, except that no anion exchange chromatography was carried out. The preparation was finally dialysed into phosphate buffered saline, 5% glycerol, 0.01% sodium azide and 0.5mM EDTA pH7.4.

The results from assays of material processed in this way indicated that some activity was obtained. The level of activity obtained in this way was too low to do any detailed studies on, so the resultant dialysate was purified by anion exchange chromatography (Figure 11). This process resulted in the isolation of significant NIP-cap-BSA binding activity over that of background binding to BSA (Figure 11). The assay of the fractions for the level of Ig μ , expressed as B1-8 IgM equivalents demonstrated two peaks of activity. This was not found to correlate with full length Ig μ by Western blotting. The first peak observed may represent a fragment of Ig μ . The separation of NIP-CAP-BSA binding activity from the majority of full length Ig μ and protein indicates that the hapten binding activity is contained within a particular molecular species formed at low efficiency.

The processing of insoluble material obtained from Ig μ expression in *E. Coli* produced a similar IgM protein profile but without NIP-cap-BSA binding activity. This demonstrates that the activity recovered was a property of the combined immunoglobulin expression, not of some *E. Coli* factor, or of the Ig μ heavy chain alone.

Further studies of the characteristics of the hapten (NIP-cap-BSA) binding were carried out. As samples were diluted they showed less binding to hapten in a very similar way to the original antibody (Figure 12). Free hapten was found to inhibit most of the binding activity in both undiluted and diluted samples. Using B1-8 antibody as a standard for both IgM and hapten binding, the specific activity of the assembled antibody was calculated to be 1.4×10^4 gm/gm of IgM equivalents. This value demonstrates the inefficient recovery of activity, but possibly represents an underestimate of the specific activity due to an overestimate of full-length Ig μ in these fractions, as described above.

Heteroclitic Nature of Recombined Antibody

Detailed specificity of binding to NIP-cap-BSA was investigated by comparing the assembled antibodies with B1-8 IgM in the presence of free NIP-cap and NP-cap (Figure 13). Both B1-8 IgM and the assembled antibodies showed that higher NP-cap than NIP-cap concentrations were required to inhibit NIP-cap-BSA binding.

The heteroclitic nature is demonstrated by the molar ratio of NIP to NP at 50% inhibition. The concentrations of NIP and NP at 50% inhibition (I50) were found to be similar for both B1-8 and the assembled antibodies as shown in Table 2. Also the NP I50/NIP I50 ratios were similar (Table 2).

Table 2. Hapten concentration at 50% inhibition (I50) of binding of antibodies to NIP-cap-BSA solid phase.

SD = Standard Deviation.

	<u>NIP I50 μM</u>	<u>NP I50 μM</u>	<u>NP I50 NIP I50</u>
B1-8 IgM	0.13 (SD, 0.05)	3.7 (SD, 2.9)	29
Fraction 26	0.34 (SD, 0.09)	1.9 (SD, 0.4)	6
Fraction 27 and 28	0.11 (SD, 0.02)	1.1 (SD, 0.3)	10
Purified μ and λ	0.4	0.84	22

It has thus been shown that not only is it possible to express μ and λ proteins having intact variable domains, but also that it is possible to express both μ and λ proteins on compatible plasmids in the same cell. This latter has not hitherto been disclosed or even suggested. Moreover, it proved possible to derive functional Ig molecules from *E. Coli* cells which expressed both μ and λ proteins.

It has thus been shown that the process of the present invention produces a functional Ig molecule by recombinant DNA techniques.

Construction of Expression Plasmids for Yeast

The plasmids used for expression in yeast (*Saccharomyces cerevisiae*) are based on pBR322 and the yeast 2 micron plasmid. These plasmids are the subject of copending European Patent Application EP 0 073 635, the disclosure of which is incorporated hereby reference. The yeast phosphoglycerate kinase gene (PGK) provides the 5' sequence necessary for initiation of transcription and insertion of genes can be made at the unique BglIII site in the plasmid pMA 3013 described in the above mentioned European patent application. This plasmid has now been renamed pMA91 as is so referred to hereinafter.

a) met- μ (μ)

BglIII and BclI Produce compatible 5'-GATC ends. The met- μ gene from pNP2 was excised on a partial BclI fragment and ligated into the unique BglIII site of pMA91.

b) Pre- μ (μ)

The plasmid pCT54 μ containing the full length pre- μ cDNA was digested with Hind III. This cuts at the 3' side of the μ cDNA. This Hind III site was changed to a BclI site by incubation with T₄ DNA polymerase in the presence of all 4 nucleotides. The linker R107 of sequence TTTTGATCAAAA, which contains an internal BclI was ligated to the DNA obtained above. After ligation one aliquot of the resultant DNA was digested with BclI and AccI and the μ AccI-BclI fragment was isolated from the resultant mixture by gel electrophoresis. A separate aliquot of the resultant DNA was digested with MboII and ligated with chemically synthesised oligonucleotide linkers R121 and R112 which have cohesive BclI and MboII ends.

R121 5' GATCAATGGGATGGAGCTGT 3'

R112 5' TTACCCTACCTCGAC 3'

The ligation reaction was terminated and the resultant DNA digested with AccI to give a μ MboII-AccI fragment which was isolated from the resultant mixture by gel electrophoresis on a 5% polyacrylamide gel.

The two μ fragments obtained above were ligated together and then digested with BclI to eliminate unwanted ligation products after which the resultant pre- μ DNA was ligated to BglIII cut plasmid pMA91.

The resultant ligation mix was used to transform *E.Coli* strain HB101 to ampicillin resistance. A colony of the transformed bacteria was isolated and was found to contain a plasmid exhibiting the predicted enzyme digestion pattern. The 5' end of the inserted DNA from this plasmid was sequenced and shown to have the anticipated sequence for a pre- μ cDNA.

c) met-lambda (λ)

The met- λ DNA was cloned into the BglIII site of plasmid pMA91. Plasmid pCT54 Clone 1 was cut with Hind III to the 3' side of the met- λ cDNA and this cleavage site was altered to a BclI site in a similar manner as previously described for pNP2. Following this the plasmid DNA was cut 5' of the λ gene at the BclI site and the resultant λ DNA was cloned into the BglIII site of pMA91.

d) Pre-lambda (λ)

Pre- λ cDNA was reconstructed into pMA 91 as follows. The plasmid pCT54 was digested with BclI and Hind III and the resultant vector DNA was isolated by gel electrophoresis and ligated together with two synthetic oligonucleotides, R162 and R163, and the two fragments from plasmid pAT λ 1-15.

R162 5' GATCAATGGCCTGGATT 3'

R163 5' GTGAAATCCAGGCCATT 3'

These fragments were produced by digesting pAT λ 1-15 with FokI and Hind III and isolating the 5' 300 base pair FokI fragment and the 3' 600 base pair FokI-Hind III fragment by gel electrophoresis. The resultant ligation mixture was used to transform *E.Coli* strain HB101 to ampicillin resistance and a colony

of bacteria containing a plasmid having the predicted correct restriction enzyme digestion pattern was isolated. The 5' end of the inserted DNA was sequenced and found to have the anticipated sequence of a pre- λ cDNA. This plasmid was digested with Hind III and the resultant Hind III cleaved ends converted to a BclI site by blunting with T₄ polymerase and ligating with the linker R107. The resultant plasmid was then digested with BclI and the pre- λ cDNA isolated on a BclI fragment by gel electrophoresis. This BclI fragment was then ligated to BglI cut pMA91 to give a pMA 91 plasmid containing the full length pre- λ cDNA.

Expression of Immunoglobulin Genes in Yeast

The pMA91 derivative plasmids containing the pre- λ and pre- μ genes as prepared above were used to transform *Saccharomyces cerevisiae* yeast host organisms and the pre- λ and pre- μ genes were expressed by the transformed cells.

Saccharomyces cerevisiae strain MD46 when transformed with pMA91 pre- λ or pMA91 pre- μ gave rise to colonies which expressed immunoreactive proteins as revealed on Western blots. Yeast cells containing plasmid pMA91 pre- λ produced a protein which reacted with anti- λ antiserum and co-migrated on polyacrylamide gels with bacterially synthesised mature λ . Similarly yeast cells containing pMA 91 pre- μ produced a protein which reacted with anti- μ antiserum and co-migrated on polyacrylamide gels with bacterially synthesised mature μ protein. These observations indicate that both pre- λ and pre- μ are being processed to the corresponding mature proteins within the yeast host cell environment.

Additionally the μ protein product (but not the λ protein) was shown to be glycosylated. Cells were incubated in the presence or absence of tunicamycin at a concentration of 15 μ g/ml. This compound specifically arrests the N-linked glycosylation of proteins. Cell extracts derived from cells incubated in the absence of tunicamycin showed higher molecular weight bands (as revealed by Western blotting) whilst extracts from cells incubated in the presence of tunicamycin showed no such higher molecular weight bands.

When cells were lysed with glass beads and the soluble and insoluble fractions examined it was found that the μ and λ proteins were exclusively in the insoluble fractions, as determined by Western blotting.

Secretion

After incubation transformed yeast cells were spun down and 1 ml volumes of supernatant were removed and passed through BSA-coated 0.2 μ m Millipore filters to remove any remaining cellular material. ELISA assays for μ and λ protein were then carried out on the filtered supernatants. Only supernatants from pMA91 pre- λ harbouring cells showed detectable levels of immunoglobulin protein. Increased amounts of λ protein was detected in the supernatant when cells were grown in minimal medium to OD₆₆₀ = 0.2 and then spun down and resuspended in YPAB and harvested at OD₆₆₀ = 1.0-1.5. Thus the λ protein but not the μ protein is secreted from yeast cells.

Intracellular Location

After incubation cells containing pMA91 pre- μ were converted to spheroplasts and fixed in 5% acetic acid/95% ethanol (v/v) and incubated with fluorescein conjugated goat anti-mouse μ . The fixed cells were examined by fluorescent microscopy and the μ protein was found to be localised in the periphery of the spheroplasts and especially in vacuoles.

Expression of both μ and λ Proteins in the Same Cells

In order to express both genes in the same host cells it was necessary to provide compatible plasmids for use in transformation. The pre- λ gene was excised from pMA91 pre- λ on a Hind III fragment and inserted into the Hind III site of plasmid pLG89 (Griti L, and Davies J, Gene, 25 179-188, 1983). This plasmid contains a ura3 marker which can be used as a positive selection for transformed host cells. A convenient ura3⁻ host organism is *S. cerevisiae* strain X4003-5B. Both plasmids, pMA91 pre- μ and pLG89 pre- λ , were transformed into this strain, and colonies were grown which contained together both the ura3 and leu2 markers. After incubation of the transformed cells both λ and μ proteins together were detected in the same cultures of X4003-5B using ELISA techniques. Levels of expression were comparable to those obtained for the individual genes in MD46 cells and for the λ gene alone in X4003-5B.

In addition, to check for assembly of the λ and μ proteins *in vivo* the following procedure was followed.

After the growth to an $OD_{550} \approx 1$, the transformed X4003-5B cells were spun down and resuspended in buffer, either 5 mM borate buffer at pH 8.0 or phosphate buffer at pH 5.8 either with or without detergent (e.g. 0.5% Triton X). The suspended cells were then lysed by vortexing with glass beads in buffer as above and the insoluble material was spun down. After centrifugation aliquots of the supernatant were assayed in a NIP binding assay (as described previously for assembled E.Coli μ and λ proteins). Specific antigen binding activity was detected in the supernatant and this activity was shown to be specifically competed out by free NIP (the specific antigen). These results indicated that μ and λ protein were expressed within the X4003-5B cells and assembled into functional immunoglobulin molecules *in vivo*.

In the above experiments the culture medium used for incubating the transformed cells was yeast minimal medium (containing, per litre, 6.7 g DIFCO yeast nitrogen base without amino acids, 10 g glucose, and 200 mg each of histidine, tryptophan, methionine and adenine). When the λ protein alone was being expressed the medium contained in addition 200 mg/l of leucine, and when the μ protein alone was being expressed the medium contained in addition 200 mg/l of uracil.

Claims

1. A process for producing a heterologous Ig molecule or an immunologically functional Ig fragment in a single host cell, which comprises transforming the host cell with separate DNA sequences respectively encoding polypeptide chains comprising at least the variable domains of the Ig heavy and light chains and expressing each of said polypeptide chains separately in said transformed single host cell.
2. The process of claim 1, wherein each DNA sequence is inserted individually into a vector and the individual vectors are used to transform the single host cell.
3. The process of claim 1, wherein the two DNA sequences are inserted into separate parts of a vector which is used to transform the single host cell.
4. The process of claim 2 or claim 3, wherein the or each vector is a plasmid.
5. The process of any one of claims 1 to 4, wherein the host cell is a eukaryotic host cell.
6. The process of claim 5, wherein the host cell is a yeast.
7. The process of claim 6, wherein the host cell is *S. cerevisiae*.
8. The process of claim 5, wherein the host cell is a mammalian cell.
9. The process of any one of claims 1 to 8, wherein the polypeptide chains are secreted by the host cell.
10. The process of any one of claims 1 to 9, for producing an Ig molecule or fragment having at least one constant domain, wherein the or each constant domain is derived from the same source as the variable domain to which it is attached.
11. The process of any one of claims 1 to 9, for producing an Ig molecule or fragment having at least one constant domain, wherein the or each constant domain is derived from a source different from that from which the variable domain to which it is attached is derived.
12. The process of any one of claims 1 to 11, wherein the DNA sequences code for complete Ig heavy and light chains.
13. The process of any one of claims 1 to 12, wherein the DNA sequences code for only one constant domain.
14. The process of any one of claims 1 to 13, wherein at least one of the DNA sequences encodes at least the variable region of an Ig heavy or light chain and also a cytotoxic, enzymic or structural polypeptide moiety whereby the host cell is transformed so as to produce an Ig molecule or fragment having attached thereto said cytotoxic, enzymic or structural polypeptide moiety.

15. The process of any one of claims 1 to 14, wherein the DNA coding sequences for the Ig molecule or fragment are derived from one or more monoclonal antibody-producing hybridomas.
16. A vector for use in the process of any one of claims 1 to 15, comprising separate DNA sequences
5 respectively encoding polypeptide chains comprising at least the variable domains of the Ig heavy and light chains.
17. A vector according to claim 16, which is a plasmid.
- 10 18. A host cell transformed with the vector of claim 16 or claim 17.
19. A host cell transformed with two separate vectors, the first vector comprising a DNA sequence encoding a polypeptide chain comprising at least the variable domain of the Ig heavy chain, and the second vector comprising a DNA sequence encoding a polypeptide chain comprising at least the
15 variable domain of the Ig light chain.

Patentansprüche

1. Verfahren zur Herstellung eines heterologen Ig-Moleküls oder eines immunologisch funktionellen Ig-Fragments in einer einzigen Wirtszelle umfassend das Transformieren der Wirtszelle mit getrennten DNA-Sequenzen, die für die entsprechenden Polypeptidketten kodieren, welche zumindest die variablen Domänen der schweren und leichten Ig-Ketten umfassen, und das getrennte Expressieren der Polypeptidketten in der transformierten einzigen Wirtszelle.
2. Verfahren nach Anspruch 1, bei dem jede DNA-Sequenz individuell in einen Vektor insertiert wird und die individuellen Vektoren zur Transformation der einzigen Wirtszelle verwendet werden.
3. Verfahren nach Anspruch 1, bei dem die beiden DNA-Sequenzen in getrennte Bereiche eines Vektors insertiert werden, welcher zur Transformation der einzigen Wirtszelle verwendet wird.
- 30 4. Verfahren nach Anspruch 2 oder 3, bei dem der oder jeder Vektor ein Plasmid ist.
5. Verfahren nach einem der Ansprüche 1 bis 4, bei dem die Wirtszelle eine eukaryontische Wirtszelle ist.
- 35 6. Verfahren nach Anspruch 5, bei dem die Wirtszelle eine Hefe ist.
7. Verfahren nach Anspruch 6, bei dem die Wirtszelle S. cerevisiae ist.
8. Verfahren nach Anspruch 5, bei dem die Wirtszelle eine Säugerzelle ist.
- 40 9. Verfahren nach einem der Ansprüche 1 bis 8, bei dem die Polypeptidketten von der Wirtszelle exprimiert und sekretiert werden.
10. Verfahren nach einem der Ansprüche 1 bis 9 zur Herstellung eines Ig-Moleküls oder -Fragments mit
45 zumindest einer konstanten Domäne, wobei sich die oder jede konstante Domäne von derselben Quelle wie die variable Domäne ableitet, an die sie angeheftet ist.
11. Verfahren nach einem der Ansprüche 1 bis 10 zur Herstellung eines Ig-Moleküls oder -Fragments mit
50 zumindest einer konstanten Domäne, wobei die oder jede konstante Domäne aus einer anderen Quelle als derjenigen stammt, aus der sich die variable Domäne ableitet, an welche sie angeheftet ist.
12. Verfahren nach einem der Ansprüche 1 bis 11, bei dem die DNA-Sequenzen für vollständige schwere und leichte Ig-Ketten kodieren.
- 55 13. Verfahren nach einem der Ansprüche 1 bis 12, bei dem die DNA-Sequenzen für lediglich eine konstante Domäne kodieren.

14. Verfahren nach einem der Ansprüche 1 bis 10, bei dem die DNA-Sequenzen für eine schwere oder leichte Ig-Kette oder ein Fragment davon mit einer vollständigen variablen Domäne kodieren.
15. Verfahren nach einem der Ansprüche 1 bis 14, bei dem zumindest eine der DNA-Sequenzen zumindest für die variable Region einer schweren oder leichten Ig-Kette und ferner für eine cytotoxische, enzymatische oder strukturelle Polypeptidkomponente kodiert, wodurch die Wirtszelle dahingehend transformiert wird, daß sie ein Ig-Molekül oder -Fragment bildet, an welches die cytotoxische, enzymatische oder strukturelle Polypeptidkomponente angeheftet ist.
16. Verfahren nach einem der Ansprüche 1 bis 15, bei dem die für das Ig-Molekül oder -Fragment kodierenden DNA-Sequenzen aus einem oder mehreren monoklonale Antikörper bildenden Hybridomzellen stammen.
17. Vektor zur Verwendung in dem Verfahren gemäß einem der Ansprüche 1 bis 16, welcher die für die entsprechenden Polypeptidketten, welche zumindest die variablen Domänen der schweren und leichten Ig-Ketten umfassen, kodierenden DNA-Sequenzen getrennt umfaßt.
18. Vektor nach Anspruch 17, welcher ein Plasmid ist.
19. Eine mit dem Vektor gemäß Anspruch 17 oder Anspruch 18 transformierte Wirtszelle.
20. Eine mit zwei getrennten Vektoren transformierte Wirtszelle, wobei der erste Vektor eine DNA-Sequenz umfaßt, die für eine Polypeptidkette kodiert, welche zumindest die variable Domäne der schweren Ig-Kette umfaßt, und der zweite Vektor eine DNA-Sequenz umfaßt, die für eine Polypeptidkette kodiert, welche zumindest die variable Domäne der leichten Ig-Kette umfaßt.

Revendications

1. Procédé pour la production d'une molécule d'Ig hétérologue ou d'un fragment d'Ig immunologiquement fonctionnel dans une cellule hôte unique, qui comprend la transformation de la cellule hôte avec des séquences d'ADN séparées codant respectivement pour des chaînes polypeptidiques comprenant au moins les domaines variables des chaînes lourdes et légères d'Ig et exprimant chacune des dites chaînes polypeptidiques séparément dans ladite cellule hôte unique transformée.
2. Procédé selon la revendication 1, dans lequel chaque séquence d'ADN est insérée individuellement dans un vecteur et les vecteurs individuels sont utilisés pour transformer la cellule hôte unique.
3. Procédé selon la revendication 1, dans lequel les deux séquences d'ADN sont insérées dans des parties séparées d'un vecteur qui est utilisé pour transformer la cellule hôte unique.
4. Procédé selon l'une des revendications 2 ou 3, dans lequel le ou chaque vecteur est un plasmide.
5. Procédé selon l'une quelconque des revendications 1 à 4, dans lequel la cellule hôte est une cellule hôte eucaryote.
6. Procédé selon la revendication 5, dans lequel la cellule hôte est une levure.
7. Procédé selon la revendication 6, dans lequel la cellule hôte est S. cerevisiae.
8. Procédé selon la revendication 5, dans lequel la cellule hôte est une cellule de mammifère.
9. Procédé selon l'une quelconque des revendications 1 à 8, dans lequel les chaînes polypeptidiques sont exprimées et sécrétées par la cellule hôte.
10. Procédé selon l'une quelconque des revendications 1 à 9, pour la production d'une molécule ou d'un fragment d'Ig ayant au moins un domaine constant, dans lequel le ou chaque domaine constant est dérivé de la même source que le domaine variable auquel il est fixé.

11. Procédé selon l'une quelconque des revendications 1 à 10, pour la production d'une molécule ou d'un fragment d'Ig ayant au moins un domaine constant, dans lequel le ou chaque domaine constant est dérivé d'une source différente de celle de laquelle le domaine variable auquel il est attaché est dérivé.
- 5 12. Procédé selon l'une quelconque des revendications 1 à 11, dans lequel les séquences d'ADN codent pour des chaînes lourdes et légères d'Ig complètes.
13. Procédé selon l'une quelconque des revendications 1 à 12, dans lequel les séquences d'ADN codent pour seulement un domaine constant.
- 10 14. Procédé selon l'une quelconque des revendications 1 à 10, dans lequel les séquences d'ADN codent pour une chaîne lourde ou légère d'Ig ou un fragment de celle-ci ayant un domaine variable complet.
- 15 15. Procédé selon l'une quelconque des revendications 1 à 14, dans lequel au moins une des séquences d'ADN code pour au moins la région variable d'une chaîne lourde ou légère d'Ig et également une unité polypeptidique cytotoxique, enzymatique ou structurale, la cellule hôte étant ainsi transformée de façon à produire une molécule ou un fragment d'Ig auquel est fixée ladite unité polypeptidique cytotoxique, enzymatique ou structurale.
- 20 16. Procédé selon l'une quelconque des revendications 1 à 15, dans lequel les séquences codantes d'ADN pour la molécule ou le fragment d'Ig sont dérivées d'un ou plusieurs hybridomes monoclonaux produisant des anticorps.
- 25 17. Vecteur pour utilisation dans le procédé selon l'une quelconque des revendications 1 à 16, comprenant des séquences d'ADN séparées codant respectivement pour des chaînes polypeptidiques comprenant au moins les domaines variables des chaînes lourdes et légères d'Ig.
18. Vecteur selon la revendication 17, qui est un plasmide.
- 30 19. Cellule hôte transformée avec le vecteur selon l'une des revendications 17 ou 18.
20. Cellule hôte transformée avec deux vecteurs séparés, le premier vecteur comprenant une séquence d'ADN codant pour une chaîne polypeptidique comprenant au moins le domaine variable de la chaîne lourde d'Ig, et le second vecteur comprenant une séquence d'ADN codant pour une chaîne polypeptidique comprenant au moins le domaine variable de la chaîne légère d'Ig.
- 35

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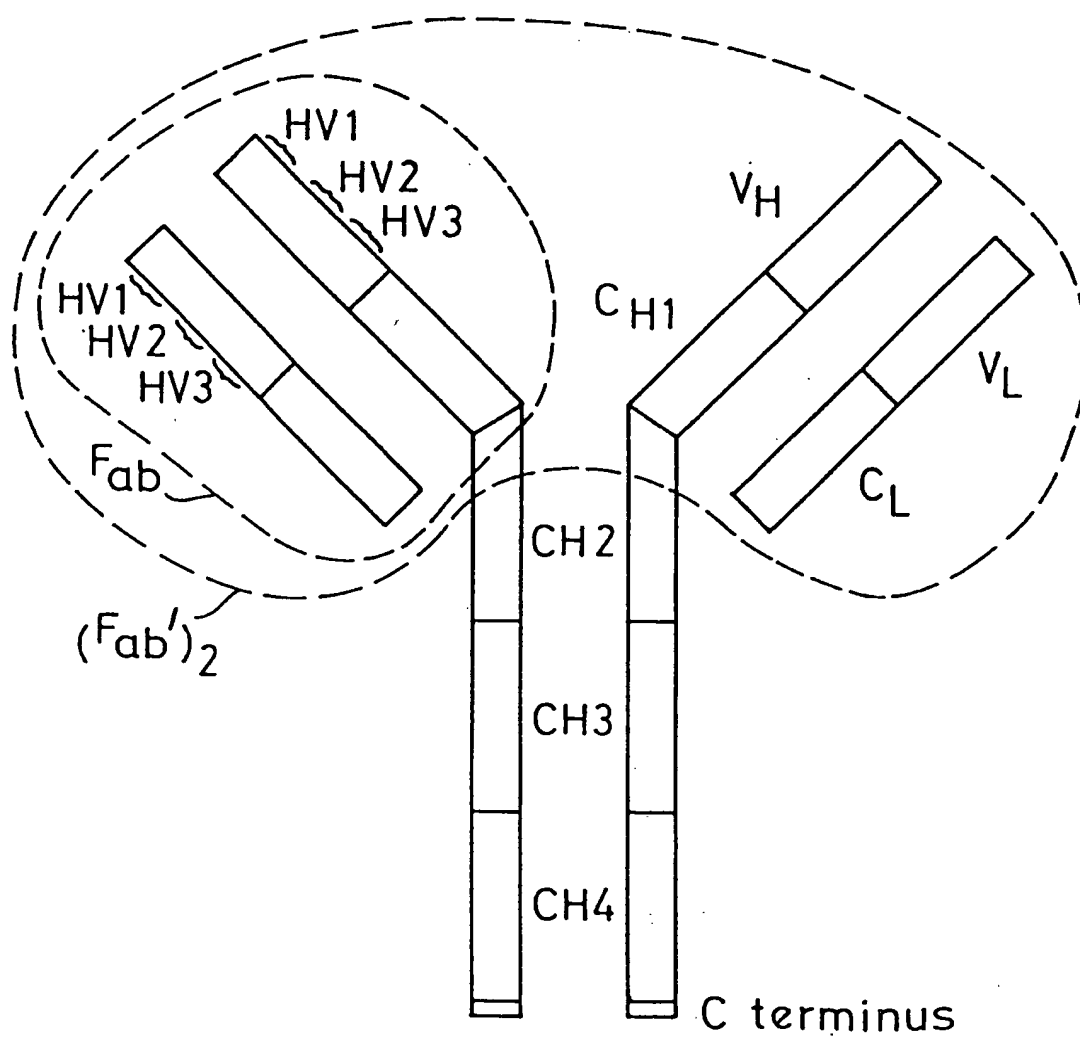
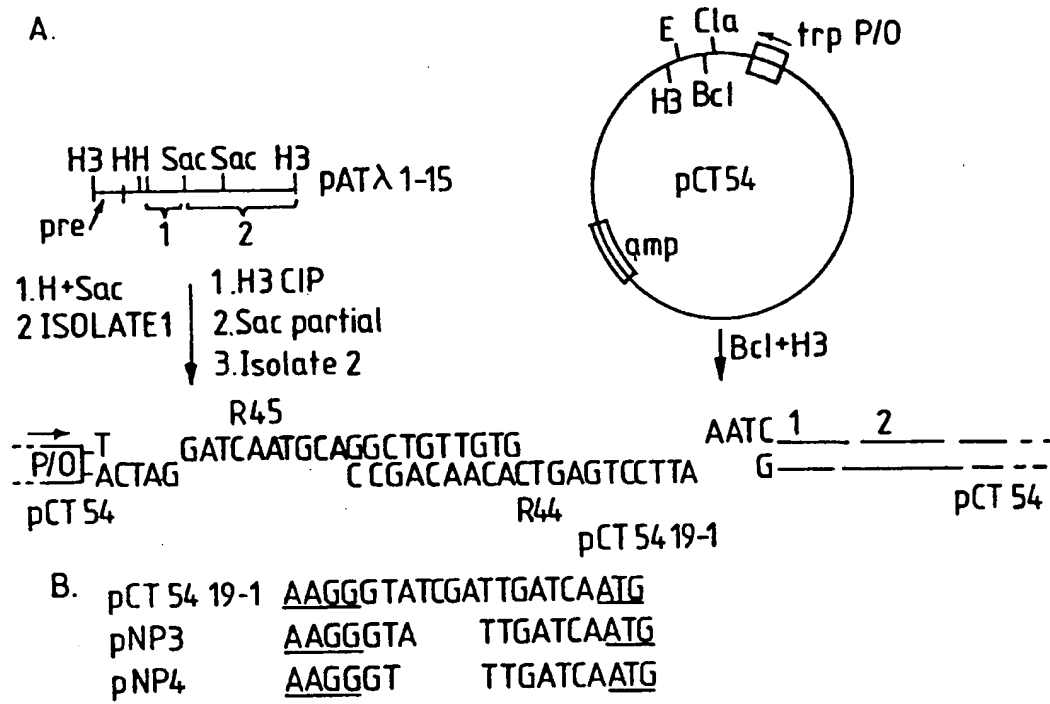


FIG. 1



Legend.

E = E corRI

H = HinfI

H3= Hind III

FIG. 2

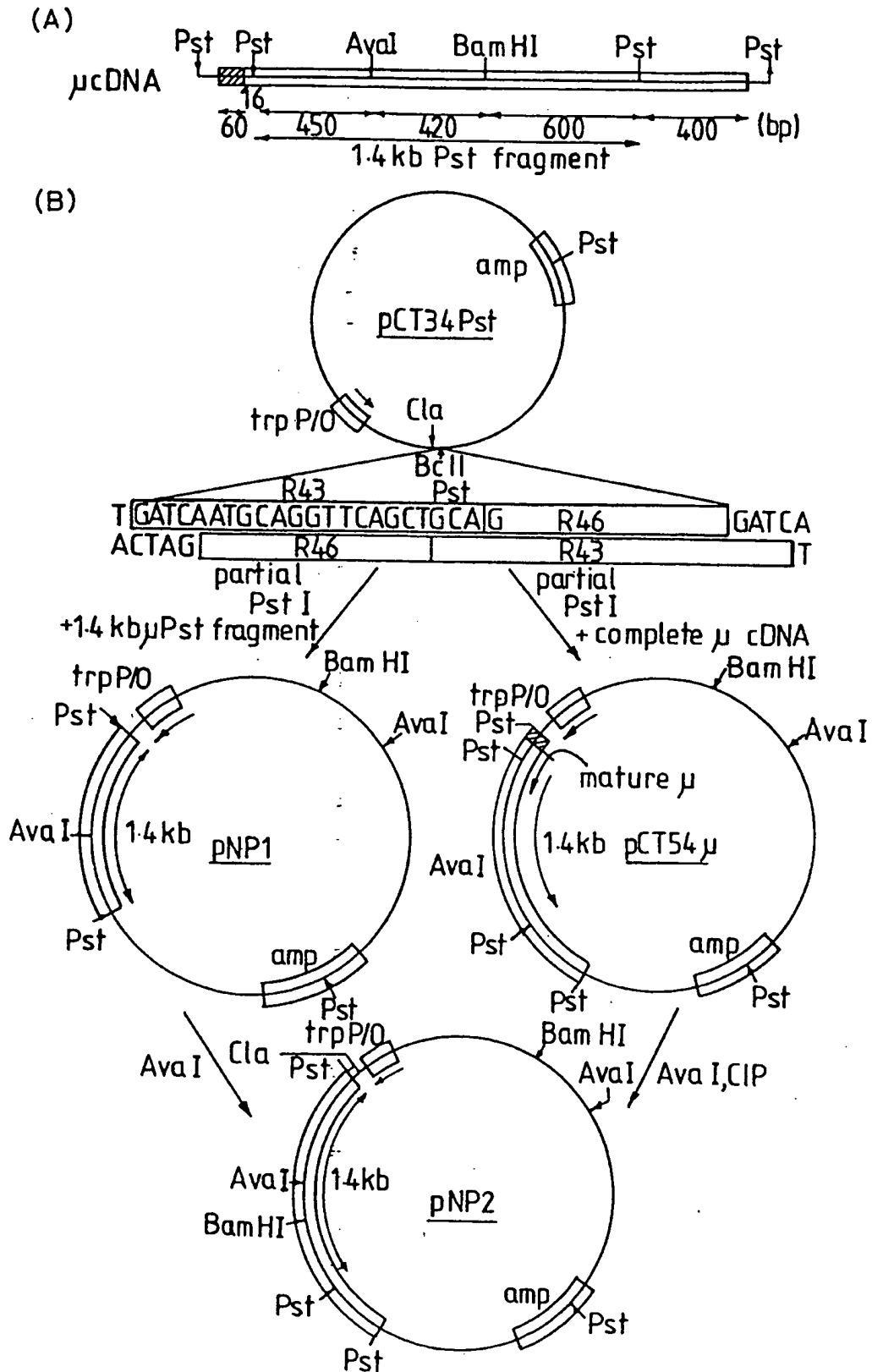
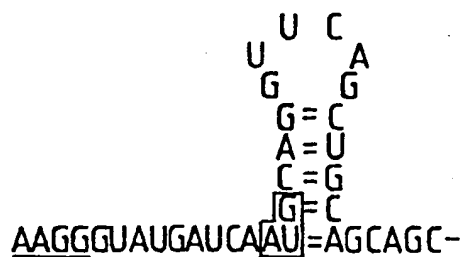


FIG. 3

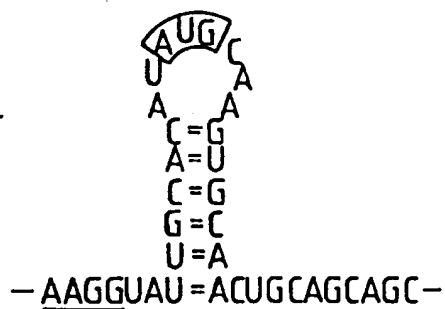
Possible 2° structures of μ mRNAs

5'→3'

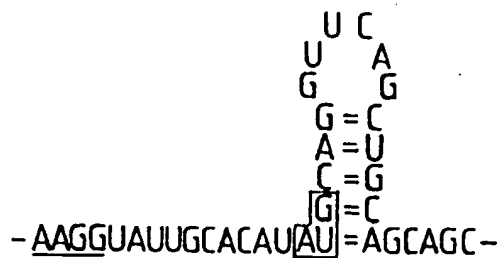
pNP9.


 $\Delta G = 7.6 \text{ K.cal.}$
(1.0 ru)

pNP11.


 $\Delta G = 7.8 \text{ K cal.}$
(6.9 ru)

pNP12


 $\Delta G = 7.6 \text{ K cal.}$
(1.8 ru)

pNP14.



(101.2 ru)

FIG. 4

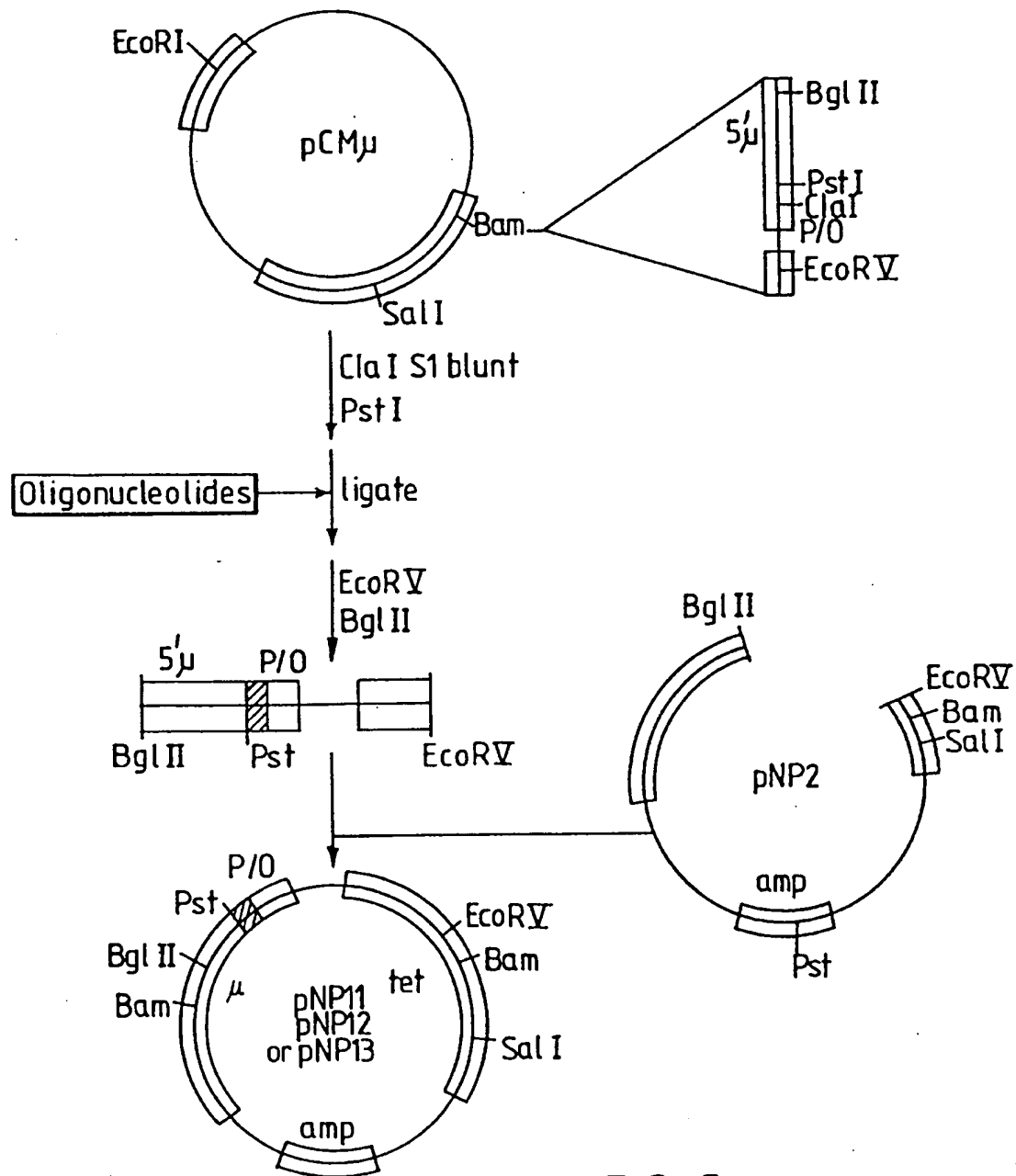


FIG. 5



FIG. 6

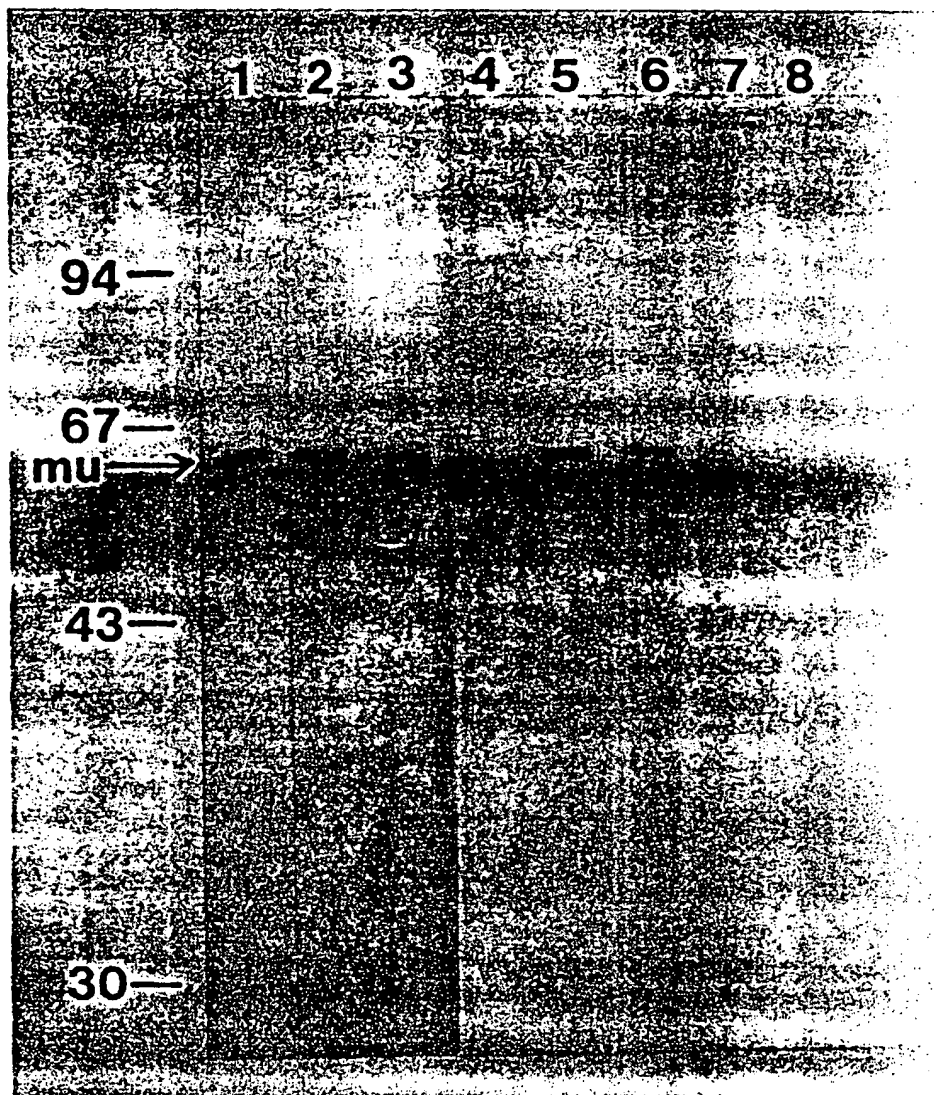


FIG. 7

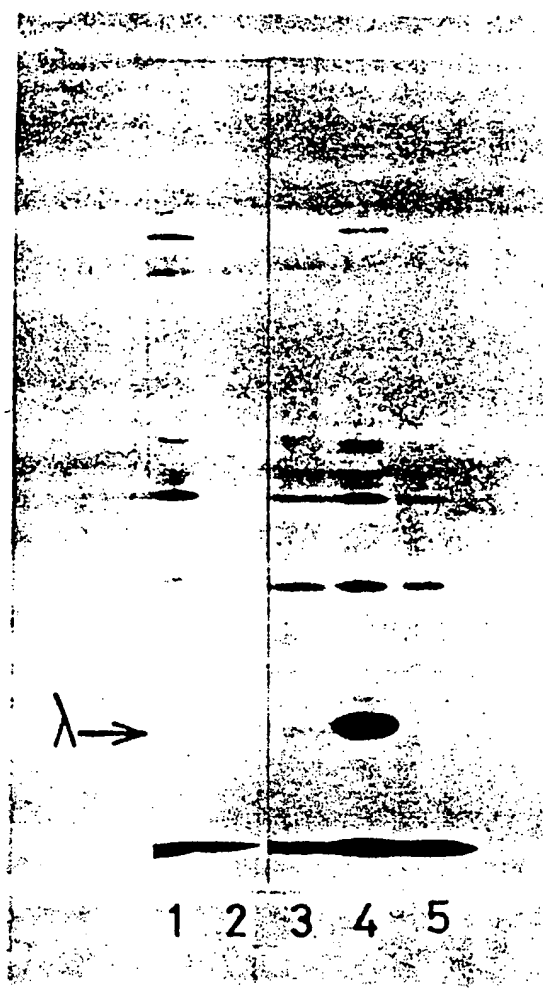


FIG. 8

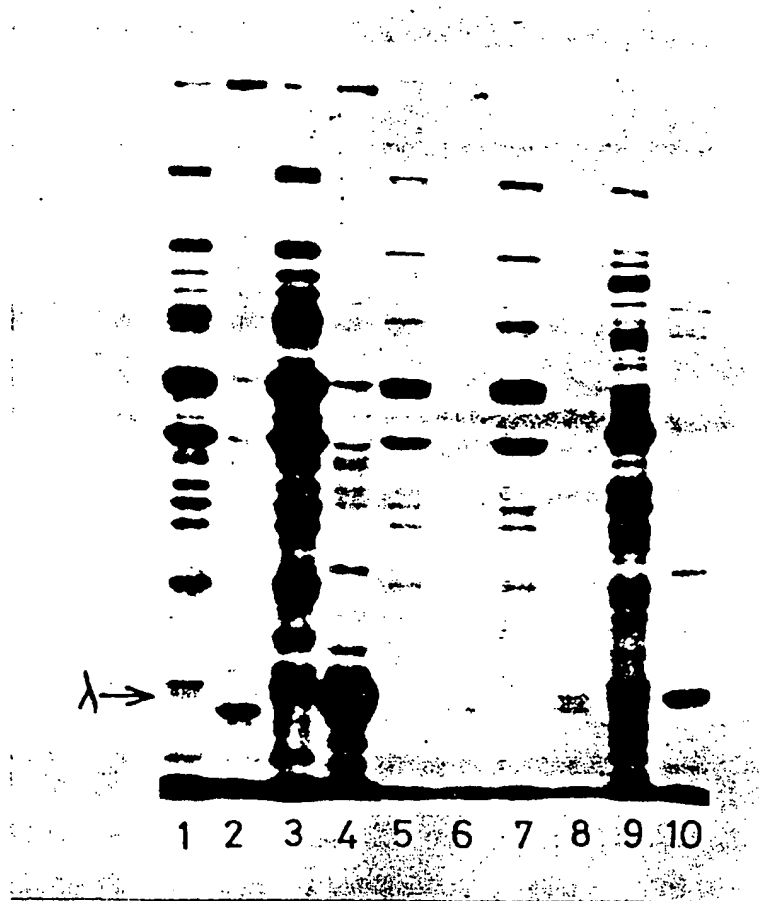


FIG. 9

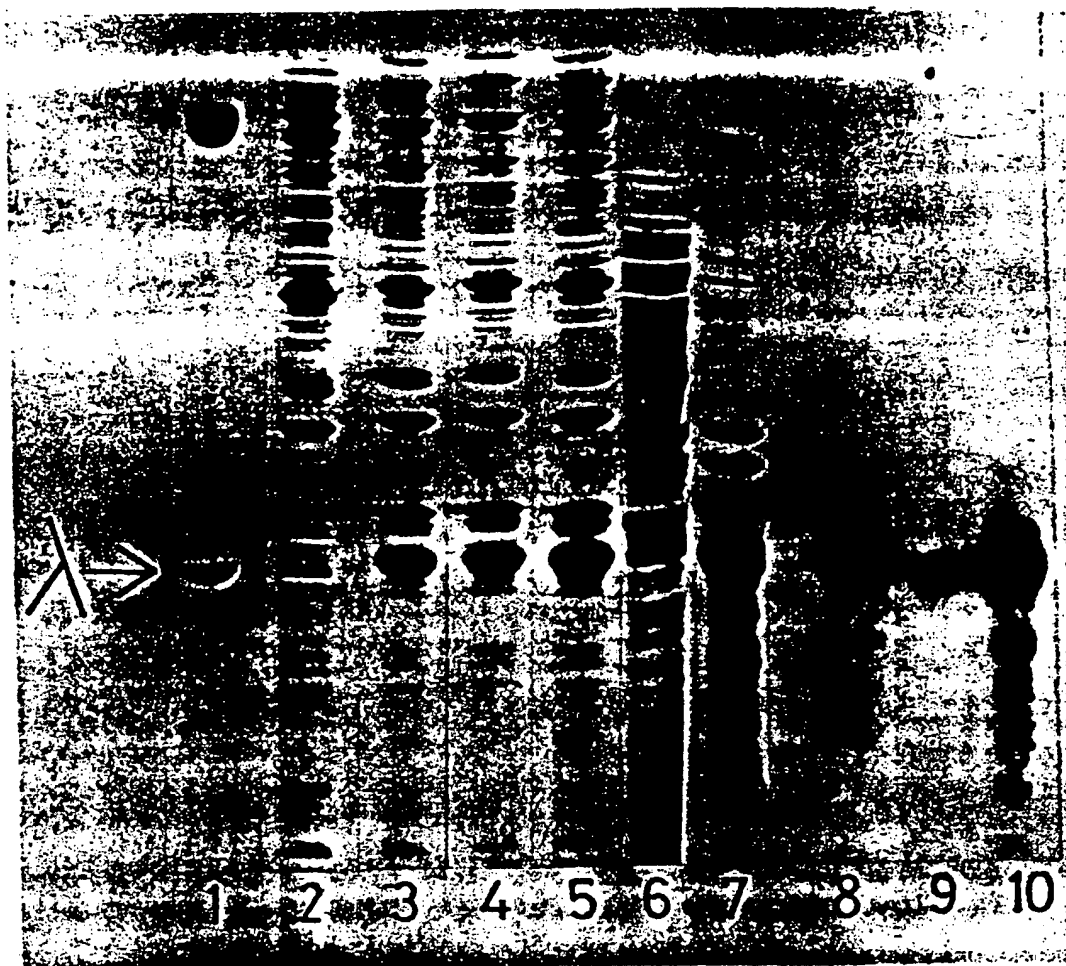


FIG. 10

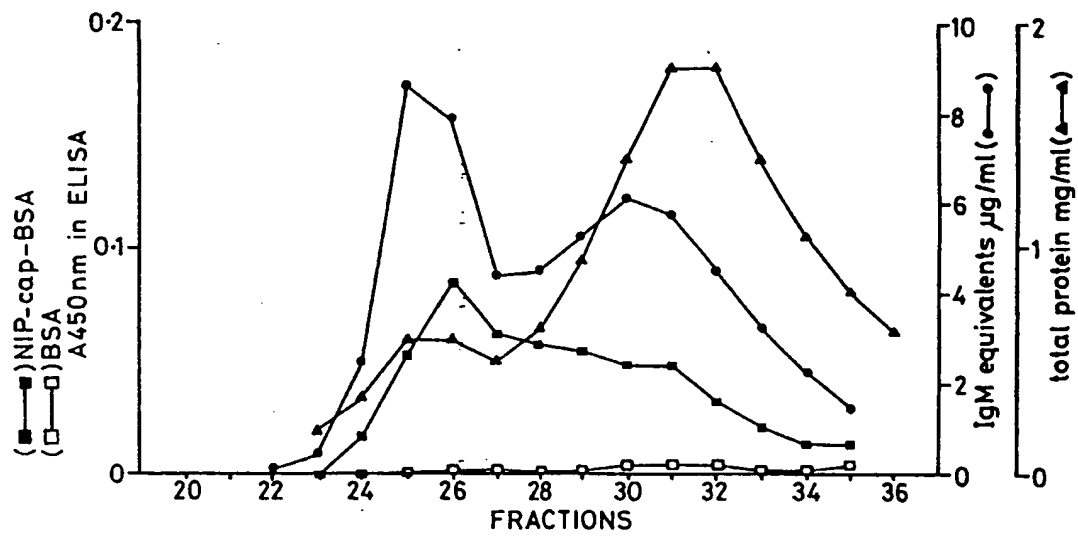


FIG. 11

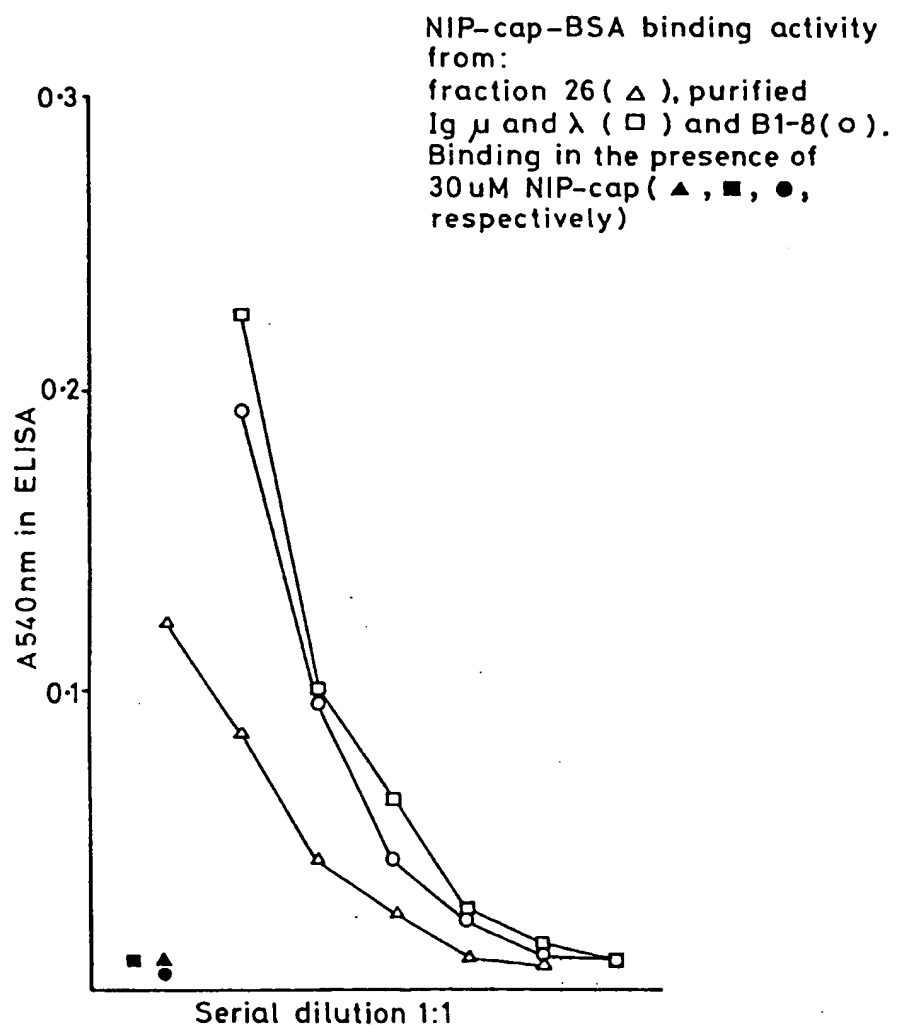
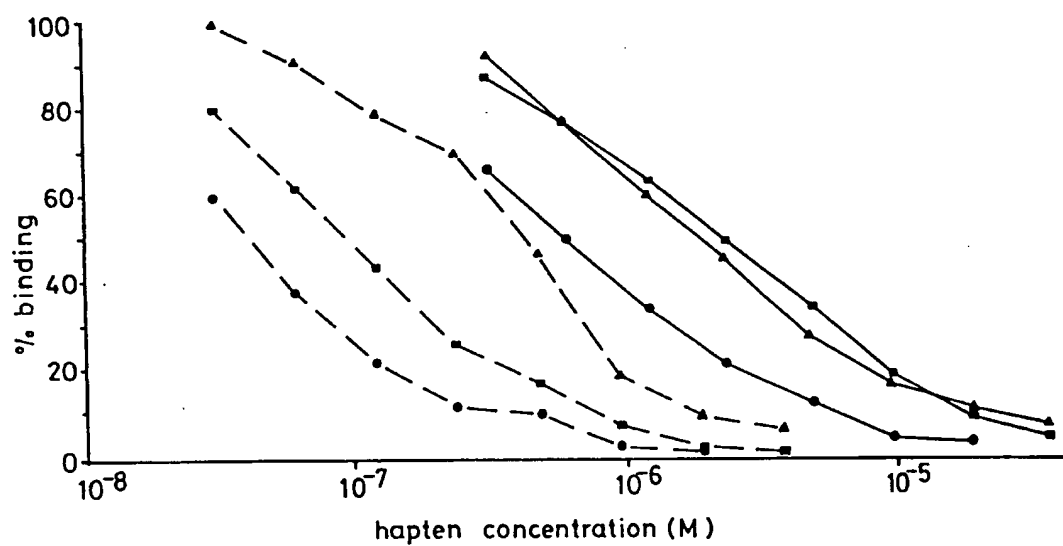


FIG. 12



Binding of antibodies to NIP-cap BSA
 B1-8 IgM (■), fraction 26 (▲),
 purified Ig μ or λ (●), in the presence
 of free NIP-cap(----) or NIP-cap (—).

FIG. 13